

Biological Forum – An International Journal

13(4): 381-387(2021)

ISSN No. (Print): 0975-1130 ISSN No. (Online): 2249-3239

Comparative Biology of Fall Armyworm, Spodoptera frugiperda on Different Host Plants under Laboratory Conditions

P. Kranthi¹ and R. Sunitha Devi^{2*}

¹M.Sc. Scholar, Department of Entomology, College of Agriculture, Professor Jayashankar Telangana State Agricultural University, (Telangana), India. ²Associate Professor, Department of Entomology, College of Agriculture, Professor Jayashankar Telangana State Agricultural University, (Telangana), India.

> (Corresponding author: R. Sunitha Devi*) (Received 28 August 2021, Accepted 25 October, 2021) (Published by Research Trend, Website: www.researchtrend.net)

ABSTRACT: Fall Armyworm, Spodoptera frugiperda (J. E. Smith) (Lepidoptera, Noctuidae) was recently reported for the first time in India in May, 2018 as a new invasive pest of maize. A detailed record of S. frugiperda's host plants is essential to better understand the biology and ecology of this pest, conduct future studies, and develop Integrated Pest Management programmes. Therefore, present study were undertaken to study biology on three different natural host plants viz., maize, sorghum, sugarcane along with artificial diet as control under laboratory conditions during kharif, 2019 at Department of Entomology, College of Agriculture, Rajendranagar, Hyderabad. During the experiment, newly hatched larvae were fed with respective host plants and artificial diet. Duration of development from larva to adult and oviposition were evaluated. The results revealed that, significant lowest larval developmental period of 12.28 ± 0.05 days was noticed on artificial diet, while it was maximum (16.82 ± 0.06 days) on sugarcane. Similar trend was observed in pre pupal and pupal period. Significantly highest male and female adult longevity (7.59 ± 0.056 and 11.36 \pm 0.128 days, respectively) was found on sugarcane followed by sorghum (7.53 \pm 0.033 and 11.27 \pm 0.163 days, respectively) and maize (7.50 \pm 0.065 days and 10.17 \pm 0.082 days, respectively), while shortest male and female adult longevity of 6.34 ± 0.084 days and 8.78 ± 0.130 days was recorded on artificial diet. The total developmental period of males and females adults was longer on sugarcane (36.81 \pm 0.16 and 40.56 \pm 0.93 days, respectively) followed by sorghum (34.35 \pm 0.22 and 39.00 \pm 0.24 days, respectively) and maize $(33.35 \pm 0.20 \text{ and } 35.63 \pm 0.37 \text{ days, respectively})$. Shortest developmental period was observed on artificial diet $(30.33 \pm 0.1 \text{ and } 32.58 \pm 0.18 \text{ days, respectively})$. Maximum fecundity was recorded when larvae were fed with artificial diet compared to natural host plants *i.e.*, maize, sorghum and sugarcane. The number of eggs oviposited was highest on artificial diet (1846.36 \pm 16.00 eggs) followed by maize (1008.36 \pm 13.35 eggs), sorghum (686.68 \pm 4.00 eggs) and lowest number of eggs were laid on sugarcane $(544.18 \pm 5.00 \text{ eggs})$.

Keywords: Spodoptera frugiperda, biology, maize, sugarcane, sorghum, artificial diet.

INTRODUCTION

The fall armyworm, Spodoptera frugiperda (J.E. Smith) (Lepidoptera: Noctuidae), which originated in the tropical and subtropical regions of America, has been identified as an notorious polyphagous pest with high migration ability, a wide range of hosts, voracious larval feeding and high fecundity; this pest is known to cause heavy economic damage to crops and pastures worldwide (Johnson, 1987; Montezano et al., 2018; Westbrook et al., 2016). It occurs in several maize growing countries such as Brazil, Argentina, Mexico and the Unites States of America (Prowell et al., 2004; Clark et al., 2007). S. frugiperda consists of two haplotypes: corn strain and rice strain. The corn-strain haplotype mainly feeds on corn, cotton and sorghum, while the rice-strain haplotype invades rice and pastures (Dumas et al., 2015). Though S. frugiperda is a key pest of maize, due to its polyphagous nature uses

important cultivated species of poaceae as its host and has ability to reach pest status on several of them (e.g., rice, wheat, sorghum, and corn) (Luginbill, 1928; Sparks, 1979; Cruz, 1999; Capinera, 2007). FAW larvae were reported on more than 60 different species of plants, particularly graminaceous hosts, such as maize, sorghum and Bermuda grass (Mitchell, 1979). The occurrence of FAW was reported in West Africa for the first time in early 2016 (Goergen et al., 2016; Abrahams et al., 2017). In Asia, S. frugiperda was first detected in India in 2018 and later in other countries, which include Myanmar, Thailand, Yemen and Sri Lanka (Deshmukh et al., 2018; FAO, 2020). In India, occurrence of this invasive pest was reported for the first time on maize from Karnataka by Sharanabasappa et al., (2018) during the month of May, 2018. Presence of FAW was observed during regular surveillance in maize fields at the College of Agriculture, Shivamogga and neighboring districts. Its occurrence was further

Kranthi & Devi

Biological Forum – An International Journal 13(4): 381-387(2021)

confirmed in Karnataka and other states like Tamil Nadu and Telangana. Molecular diversity of fall armyworm, S. frugiperda was studied from different states of India and indicated prevalence of R-strain. India being a subtropical cultivates most of graminaceious food crops such as maize, wheat, rice, sorghum, sugarcane and many minor millets in all parts of the country. The invasive pest though prefers maize but being polyphagous could turn out to be a potential threat to food security of the country. Moreover, the planting seasons of different crops are often overlapping or continuous in different regions of India, which could provide sufficient food resources for the occurrence and migration of S. frugiperda. It is well known that plant species significantly affect the survival, fecundity and population growth of herbivorous insects (Awmack et al., 2002). The impact of plant species that slow or accelerate herbivore development should be taken into account when designing and developing integrated pest managements. Therefore, investigating the effects of most host crops on the growth, development, survival and reproduction of S. frugiperda is of great significance to make a comprehensive control strategy and predict the occurrence of the population. Jing-Fei Guo et al., (2021) studied the biology of FAW on different host plants and found that larvae fed on maize exhibited significantly higher survival than those fed on potato and tobacco. Alton et al., (1979) found that corn, peanuts, sorghum are favored hosts for larvae of FAW. Ribeiro et al., (2020) studied the biological performance of S. frugiperda on different host plants and suggested that bermudagrass is the most suitable alternative host for the development of S. frugiperda. Silva et al., (2017) studied the biology of S. frugiperda using different food sources viz., soybean, cotton, maize, wheat, and oat leaves and artificial diet as the control and reported that grasses were better hosts for S. frugiperda development. Cotton was the least preferred food, followed by soybean.

To develop effective management strategies for S. frugiperda in its new invasive habitat, basic biological and ecological knowledge of this pest on different crops are crucial requirements. Reportedly, S. frugiperda has the potential to damage 353 species of plants belonging to 76 plant families (Montezano et al., 2018). However, to our knowledge, the effects of most host plants on the biological characteristics of S. frugiperda have not been well studied in India. Further, presence of good number of R-strain population and the potentiality of this pest to become a major pest on other graminacious crop plants an attempt was made to know the biology of S. frugiperda on different graminacious plants. The results of present study could help to determine the food preferences and possible population build up of S. frugiperda in turn helps in designing the management strategy.

MATERIAL AND METHODS

The present investigation on "Comparative biology of fall armyworm, *S. frugiperda* on different host plants

under laboratory conditions" was carried out in the Department of Entomology, College of Agriculture, Rajendranagar, Hyderabad during kharif, 2019-20. Biology were studied on three different natural host plants viz., maize, sorghum, sugarcane along with artificial diet as control. Each of these plants were sown during first fortnight of July, 2019 in the red sandy loam soils in an area of 200 m^2 . These plants were sown to provide continuous supply of food for S. frugiperda larvae needed for conducting the experiment. Gap filling and thinning was done after a week of germination. Crop was kept weed free through regular hand weeding. The plots were regularly irrigated, whenever the top two to three inches of soil was found dry. All the recommended agronomic package of practice were followed for raising the crops. Special care was taken to avoid application of chemical spray to the cultivated host plants.

To study the biology of *S. frugiperda* on different host plants, mass multiplication of *S. frugiperda* was taken up on artificial diet.

Experimental details. Present investigation was carried out to study the effect of different host plants and artificial diet on the development of different stages of *S. frugiperda* under laboratory. During the experimental period the average room temperature was maintained at $25 \pm 2^{\circ}$ C and the relative humidity was maintained at 70 ± 5 per cent.

Larval feeding test. Freshly hatched, neonate larvae, (0-12 hrs. old) of S. frugiperda obtained from the laboratory that were reared on artificial diet were used for the experiment. Neonate larvae were released individually in petri plates containing fresh leaves of selected host plants viz., maize, sorghum, sugarcane along with artificial diet. The larvae were reared on host plants till pupation and adult emergence. Fresh leaves of respective host plants were brought to the laboratory in separate polythene covers. Plant leaves were cleaned in distilled water, shade dried and later sand wiched between the two layers of blotting paper for removing the water. The leaves were then cut into small discs of around 7-8 cm diameter. These leaf discs were placed in small petri plates of 9 cm diameter containing circularly cut moist filter paper, to avoid drying of leaf discs. Proper care was taken to prevent the escape of larvae by covering petri plates with tissue para film paper and the lid was tightly secured with the help of rubber band. Leftover food material along with excreta were removed daily. Fresh leaves of host plants were provided to larvae at every 24 hrs. interval. This process was continued until the larvae entered into final instar. The final instar larvae were collected and transferred into another jar containing sand for pupation. Separate jars were used for pupation of final instar larvae of S. frugiperda that were reared on different host plants. Pupae thus formed were collected and placed in small plastic jars and covered with muslin cloth for adult emergence. Each treatment was replicated six times with 10 larvae in each replication. Each petri plate was examined daily for recording the

observations on larval period, pre pupal period and pupal period.

Adult longevity and fecundity test. To study the impact of host plant on fecundity and adult longevity, a pair of freshly emerged healthy male and female adults that were reared on a particular host plant viz., maize, sorghum, sugarcane and artificial diet were selected. The adults were then released into plastic jars for mating. The jars were lined with yellow paper as substratum for egg laying and were covered with white muslin cloth that was held in position with the help of the rubber band. The adults were fed with 10 per cent honey solution soaked in cotton swab placed in plastic cup inside the jar which was replaced daily. The eggs laid on the yellow paper and white muslin cloth were collected daily till the female stopped laying eggs. The eggs were collected daily and were placed in a separate jar for hatching. The eggs were counted daily using

hand lens and were recorded. The experiment was replicated six times with ten such pairs of adults per replication. A total of sixty pairs of adults were tested for adult longevity and fecundity. Average number of eggs laid per female and percent viability of the eggs from each treatment was calculated. Observations were recorded at every 24 hrs. interval on pre oviposition period, oviposition period, post oviposition period, adult longevity of male and female, sex ratio, fecundity, incubation period etc.

RESULTS AND DISCUSSION

Comparative biology of *S. frugiperda* fed on different host plants and artificial diet as check was studied under laboratory conditions at $27 \pm 2^{\circ}$ C temperature and 65 ± 5 % RH during *kharif*, 2019-20. The results of the investigation are presented in Table 1 and Fig. 1.

Host plant	Incubation period	Total larval period	Pre pupal Period	Pupal period	Adult longevity		Total life cycle		
					Male	Female	Male	Female	Fecundity
Maize	2.40±0.037 ^b	13.43±0.05°	2.41±0.031 ^b	7.40±0.052 ^c	7.50±0.065ª	10.17±0.082 ^b	33.35±0.20°	35.63±0.37°	1009.24±13.35 ^b
Sorghum	2.47±0.021 ^b	15.30±0.07 ^b	2.45±0.022 ^b	7.88±0.031 ^b	7.53±0.033ª	11.27±0.163ª	34.35±0.22 ^b	39.00±0.24 ^b	686.68±4.00 ^c
Sugarcane	2.58±0.04ª	16.82±0.06 ^a	2.55±0.022 ^a	8.22±0.031ª	7.59±0.056ª	11.36±0.128ª	36.81±0.16 ^a	40.56±0.93ª	544.18±5.00 ^d
Artificial diet	2.28±0.031 ^c	12.28±0.05 ^d	2.33±0.021 ^c	6.94±0.067 ^d	6.34±0.084 ^b	8.78±0.130 ^c	30.33±0.13 ^d	32.58±0.18 ^d	1846.36±16.00ª
S.E ±	0.032	0.057	0.024	0.047	0.062	0.129	0.181	0.524	10.174
CD (5%)	0.094	0.170	0.072	0.139	0.184	0.380	0.534	1.540	31.689

Table 1: Comparative biology of fall armyworm, Spodoptera frugiperda on different host plants and artificial diet.

Means followed by same letters in the column did not differ significantly by DMRT (P= 0.05%)

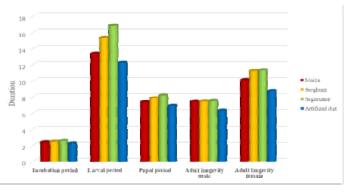


Fig. 1. Comparative biology of fall armyworm, Spodoptera frugiperda on different host plants and artificial diet.

Incubation period. The mean incubation period of S. frugiperda eggs ranged between 2.28 ± 0.031 and 2.58 \pm 0.04 days on different host plants. The incubation period was significantly less for the eggs that were laid by the females fed with artificial diet while it was more for the eggs that were laid by the females fed with sugarcane. The incubation period of eggs that were laid by females fed with sorghum and maize were on par with each other but were significantly higher compared the eggs laid by females fed with artificial diet. Similar variation in the incubation period of S. frugiperda eggs laid by females fed with different host plants was also reported by Murua et al., (2004) working on maize, Guinea grass and Bermuda grass. It was also reported that not only the nutritional variations in different host plants but also variations in the quality of nutrition in different cultivars belonging to a given host also impacts incubation period of eggs was suggested by Rosa *et al.*, (2012). According to Rosa *et al.*, (2012) the incubation period of eggs of *S. frugiperda* fed on different maize cultivars varied between 2.8 to 3.3 days. However, Azidah and Sofian-Azirun, (2006) indicated that variations in the quality of host plants did not influence the incubation period of eggs.

Total larval period. The results on total larval period of *S. frugiperda* fed on different host plants revealed that, overall larval development was significantly affected by the host plants. Data indicated that the mean larval period of *S. frugiperda* was significantly highest on sugarcane (16.82 \pm 0.06 days) followed by sorghum (15.30 \pm 0.07 days) and maize (13.43 \pm 0.05 days). Significantly, lowest mean larval duration was recorded

on artificial diet (12.28 \pm 0.05 days). It is quite evident that insect larvae consume food till their energy requirements are fulfilled. As the food consumed by the larvae has to transform into energy which helps to sustain the next active stage of the insect that is adult. The artificial diet consists of balanced nutrition viz., carbohydrates, proteins, fats including essential vitamins which generate the required quantum of energy the larva needed to transfer itself to pupa. Therefore, the larva fed with artificial diet must have acquired the necessary quantum of food and energy quickly than those fed with other plant materials. This could be the probable reason for significant short larval period of S. frugiperda fed with artificial diet. However, significant variations in the larval periods of S. frugiperda fed with maize, sorghum and sugar cane may be due to variations in the nutritional composition of these plants. Maize must be possessing the nutritional composition that helps to fulfill the muchneeded energy requirements to transfer itself to pupa in a short period compared to sorghum and sugarcane. Sorghum and sugarcane may be possessing little less nutritional composition compared to maize, that must have compelled larvae to consume smaller quantities of comparatively undesired food over extended period of feeding to derive the required quantity of nutrition. Maize has been preferred host to fall armyworm probably due to the presence of good nutritional composition that is suitable for the insect for its faster growth and to promote many parallel generations. The findings of the present investigations were in accordance with the findings of Xue-Ming et al., (2010) who have stated that the variation in nutritional of host plants composition impacts larval developmental duration. Similar results were also put forth by Farahani et al., (2011) after recording lowest larval duration of 11.98 days on mustard, 12.53 days on goosefoot, 13.10 days on soybean, 14.91 days on maize and 15.50 days on cotton by beet armyworm. FAW reared on maize exhibited the strongest performance with shorter larval developmental duration compared to potato and tobacco was also reported by Jing -Fei Guo et al., (2021). Among different host plants tested, shortest larval development period (23.8±0.3days) were reported in larvae fed with maize where as it was the longest in brinjal. (Wijerathna et al., 2021). Sá et al., (2009) reported that there is no significant differences on FAW larval development time when reared on natural hosts *i.e.*, maize, grain sorghum, Johnson grass, soybean, Brachiaria and tobacco, but it was longer for larvae reared on artificial diet. Barros et al., (2010) investigated the performance of FAW on three major crops cultivated in the Cerrado viz., soybean, corn, and cotton and millet and found that survival of FAW larvae caged on millet plants was higher than on other hosts. The FAW reared on millet also exhibited a net reproductive rate similar to that observed on corn, which was considered the best host for FAW.

Pre pupal and pupal period. Subsequent effects of host plants nutrition have been manifested in pre pupal and pupal development. The pre pupal and pupal development was also affected significantly by hosts. Kranthi & Devi

The longest mean duration of 2.55 \pm 0.022 and 8.22 \pm 0.031 days was recorded for pre pupae and pupae obtained from the larvae fed on sugarcane while the period was significantly low in larvae fed with sorghum and maize leaves which was recorded as 2.45 \pm 0.022 and 7.88 \pm 0.031 and 2.41 \pm 0.031 and 7.40 \pm 0.052 days, respectively. The shortest duration was recorded

on artificial diet (2.33 ± 0.021 and 6.94 ± 0.067 days). Larvae after gaining sufficient energy stops feeding compresses its body size and tries to convert itself into pupa. Lot of biochemical changes takes place during this process which are related to hormones and transformation of energy. Hormones play a vital role in the metamorphosis of insects and are synthesized by the nutrition the insects derive from its food. Though hormones are required in minor quantity to initiate the physiological or biochemical process in living organisms but in case of insect metamorphosis their titre is very important to promote from one stage to other. Therefore, it can be assessed that larvae fed with balanced nutrition (artificial diet) must have synthesized required quantity of hormones in a short period that helped them to convert itself from larva to pupa in lesser duration compared to those that fed on maize, sorghum or sugarcane. Similarly, pupa being a non-feeding resting stage of the insect conserves energy that was accumulated by the larva by feeding on food and transforms it to production of different parts of the adult insect which includes wings, antennae, apart from head, thorax and abdomen. The duration of pupa converting to adult also depends on rapid processing of biochemical and physiological process which require energy. If the quantum of energy in pupa is high the duration which the pupa takes itself to turn into adult will be less. Hence, the reduced pupal duration for those pupae derived from the larvae fed with artificial diet may be due to possession of high quantum of energy in them. Similar variations in the pre pupal periodof fall armyworm, S. frugiperda on different host plants was reported Rosa et al., (2012). Murúa et al., (2004) also reported variations in the pupal period of S. frugiperda fed on maize, Guinea grass and Bermuda grass. The present observations were in comparison with the studies given by Abdullah et al., (2019) who reported reduced pupal development period of S. litura fed on maize.

Adult longevity. The results presented in the Table 1, revealed that female adults lived little longer compared to males fed on different host plants and artificial diet. The reason being quite evident in all the species that females has to find a suitable location containing ample food source to lav eggs. The data also shows that the adult longevities of males and females were on par with each other except the female adults that were derived from the larvae fed with maize. However, the male and female adult longevities were significantly different from the male and female adult longevities that were derived from the larvae fed with artificial diet. Longest male life span was recorded on sugarcane (7.59 ± 0.056) days) followed by sorghum (7.53 \pm 0.033 days) and maize $(7.50 \pm 0.065 \text{ days})$ which were found to be on par with each other. Significantly shortest male life Biological Forum – An International Journal 13(4): 381-387(2021) 384

span of 6.34 ± 0.084 days was recorded on artificial diet. Similarly, longest female life span of 11.36 \pm 0.128 days was recorded on sugarcane followed by sorghum (11.27 \pm 0.163 days) which were found to be on par with each other. Significantly shortest female life span was recorded on artificial diet (8.78 \pm 0.130 days) followed by Maize (10.17 \pm 0.082 days). However, significantly short life span of adults derived from the larvae fed on artificial diet compared to those fed on different host plants may be due to their higher fecundity in short span of time. The males and females soon after their emergence mate wherein, the males donate the sperms to females once they discharge the sperms, the males perish. The females possessing good quantum of proteins produce large number of eggs within short period of time which get fertilized using the sperms donated by males. Once the females convert their energy into eggs in a short span, they get exhausted and will perish. The longer adult longevity of males and females fed with low nutritional food may be due to low energies and slow physiological processes within them leading to delayed as well as low production of eggs. Similar results were put forth by Rosa et al., (2012) who reported adult longevity of S. frugiperda varied between 14 to 32 days when reared on different maize cultivars. Murua et al., (2004) also reported adult longevity as 16.00 ± 2.8 , 17.35 ± 5.39 and 16.23 ± 4.69 days on maize, Guinea grass and Bermuda grass, respectively. Similar variability in adult longevity was also reported by Barros et al., (2010) when S. frugiperda were fed with cotton, millet, corn and soybean. FAW reared on maize exhibited longer longevity and a higher reproductive rate in adults in preference to potato or tobacco (Jing -FeiGuo et al., (2021).

Total developmental period. The total developmental period of both males as well as females reared on different host plants were significantly long compared to those reared on artificial diet. However, among different host plants S. frugiperda that were reared on sugarcane showed significant longer total developmental period of 36.81 ± 0.16 days followed by sorghum (34.35 \pm 0.22 days) and maize (33.35 u \pm 0.20 days). Significantly, shortest total development period of male as 30.33± 0.13 days was recorded on artificial diet. Similar trend was observed in respect of females wherein significant longest total developmental period was observed on sugarcane (40.56 \pm 0.93 days) followed by sorghum (39.00 \pm 0.24 days) and maize $(35.63 \pm 0.37 \text{ days})$. Shortest female total development period of 32.58± 0.18 days was recorded on artificial diet. The total developmental period depends up on the period of egg incubation, duration of larva, duration of pre pupa, pupa and adult. The reasons cited for the increase in the duration of egg, larva, pupa and adult for the insects reared on different hosts holds valid for the probable increase in the duration of total developmental period of insects. The results on variation in the total developmental period with variation in the host plant was reported by Sharma, (1994) who observed the total developmental period of S. litura to be 32.67 days on germinating seeds of soybean and 43.72 days on Biological Forum – An International Journal 13(4): 381-387(2021)

linseed. Similarly, Farahani et al., (2011) observed variation in the total developmental period of S. exigua reared on different hosts and reported that the lowest total development period on mustard with 34.12 days followed by soybean (35.15 days), goose foot (36.33 days), maize (38.42 days) and longest development period on cotton (39.94 days).

Fecundity. Number of eggs laid by S. frugiperda females fed on different host plants differed significantly. Fecundity was higher when larvae were fed with artificial diet compared to natural host plants i.e., maize, sorghum and sugarcane. The number of eggs oviposited was highest on artificial diet (1846.36 \pm 16.00 eggs) followed by maize (1009.24 \pm 13.35 eggs), sorghum (686.68 \pm 4.00) and lowest number of eggs were laid on sugarcane (544.18 \pm 5.00 eggs). Fecundity of female depends on the availability of proteins. As the artificial diet being rich in protein could be the probable reason for production of more eggs by the adults whose larvae were reared on it. The monocot grasses maize, sorghum and sugar cane being rich in carbohydrates with little protein content could be the reason for low fecundity of adults. Variation in fecundity of females that were reared on different host plants were also reported by Abdullah et al. (2019) who observed higher fecundity of S. litura fed with leaves of cabbage (2455.5), alfalfa (1750) compared to maize (1055.6). Similarly, Barros et al. (2010) reported fecundity of fall armyworm as 1144.7 \pm 132.7, 1574.1 \pm 177.6, 1604.2 \pm 353.8 and 1590.8 \pm 381.7 eggs on cotton, millet, corn and soybean, respectively. Castro and Pitre (1988) found that there is no significant difference in the fall armyworm development cycle when fed with sorghum and maize. Jing Fei Guo et al., (2021) reported that females oviposited on maize in preference to potato or tobacco. Oviposition of FAW on transgenic and conventional maize was significantly higher than that on wheat, sorghum, foxtail millet, peanut and soybean while showing no significant difference between transgenic or conventional maize (Li-mei et al., 2021). The highest oviposition $(4.1\pm1.2$ in Choice test and 3.6±0.7 in No-choice test) was observed in maize compared to other crops (Wijerathna et al., 2021). Wang et al., (2020) examined the effects of six cash crops maize, wheat, soybean, tomato, cotton and Chinese cabbage on the development, survival, fecundity of S. frugiperda and reported that the preadult stage, adult preoviposition period and total preoviposition period were shortest on maize and wheat but were longest on tomato. Fecundity was greatest on maize and wheat but smallest on tomato.

CONCLUSION

From the present study, the comparative biology of S. frugiperda is important to know the best host that support the development of S. frugiperda and can be used for mass rearing of S. frugiperda. Similarly, the information of life history parameters of S. frugiperda on different host plant species will help to make efficient strategies to control this economic pest. The comparative biology of S. frugiperda on different hosts inferred that, highest mean larval period, pre pupal and

pupal period was recorded on sugarcane while, it was lowest on artificial diet. The total developmental period of both males and females was longer on sugarcane followed by sorghum and maize, while shortest developmental period was found on artificial diet. Maximum fecundity was recorded when larvae were fed with artificial diet compared to natural host plants *i.e.*, maize, sorghum and sugarcane. The results clearly indicated that the larva fed with artificial diet showed significant shorter larval, pre pupal, pupal period and male and female adult longevity compared to larvae fed with host plants maize, sorghum and sugarcane. This could probably be due to the acquisition of necessary quantum of food and energy quickly than those fed with other plant materials. However, significant variations in the larval periods of S. frugiperda fed with maize, sorghum and sugarcane may be due to the variations in the nutritional composition of these plants. It can be concluded from the study that, among the host plants maize was the preferred host to fall armyworm. The reason for which can be attributed to the presence of good nutritional composition that was suitable for the faster growth and development of S. frugiperda and also to promote many parallel generations.

FUTURE SCOPE

The research provides idea and knowledge about the survival mechanism of *S. frugiperda* during off season of maize and potential alternative host plants of the pest. It identifies the possibility of moving of *S. frugiperda* into other graminaceous crops and thereby make awareness to the farmers about the pest, in turn to follow the suitable management practice against *S. frugiperda*.

Acknowlegement. The authors are thankful to the Professor & Head, Department of Entomology for providing all the necessary facilities during investigation for smooth conduct of experiment. Conflict of Interest. None.

REFERENCES

- Abdullah, A., Ullah, M. I., Raza, A. B. M., Arshad, M., & Afzal, M. (2019). Host Plant Selection Affects Biological Parameters in Armyworm, *Spodoptera litura* (Lepidoptera: Noctuidae). *Pakistan Journal of Zoology*, 51(6): 2117-2123.
- Abrahams, P., Bateman, M., Beale, T., Clottey, V., Cock, M., Colmenarez, Y., Corniani, N., Early, R., Day, R., Godwin, J. L., Gomez, J., Moreno, P.G., Murphy, S.T., Oppong-Mensah, B., Phiri, N., Corin, S. S., & Witt, A. (2017). Fall Armyworm: Impacts and Implications for Africa. *Outlooks on Pest Management*, 28(5): 196-201.
- Alton, N. S. (1979). Review of the biology of the fall armyworm. *Florida Entomologist*, 62(2): 82-87.
- Andrews, K. L. (1980). The whorl worm, Spodoptera frugiperda, in Central America and neighboring areas. Florida Entomologist, 63: 456-467.
- Awmack, C. S., & Leather, S. R. (2002). Host plant quality and fecundity in herbivorous insects. *Annual Review* of Entomology, 47: 817–844.

- Azidah, A. A., & Sofian-Azirun, M. (2006). Life history of Spodoptera exigua (Lepidoptera: Noctuidae) on various host plants. Bulletin of Entomological Research, 96: 613-618.
- Barros, E. M., Torres, J. B., Ruberson, J. R., & Oliveira, M. D. (2010). Development of *Spodoptera frugiperda* on different hosts and damage to reproductive structures in cotton. *Entomologia Experimentaliset Applicata*, 137(3): 237-245.
- Capinera, J. L. (2007). Fall Armyworm, Spodoptera frugiperda (J.E. Smith) (Insecta: Lepidoptera: Noctuidae). Document nr. EENY098. Department of Entomology and Nematology, UF/IFAS Extension. http://edis.ifas.ufl.edu. http://edis.ifas.ufl.edu.
- Castro, M. T., & Pitre, H. N. (1988). Development of fall armyworm, *Spodoptera frugiperda*, from Honduras and Mississippi on sorghum or corn in the laboratory. *Florida Entomologist*, 71(1): 49-56.
- Clark, P. L., Molina-Ochoa, J., Martinelli, S., Skoda, S. R., Isenhour, D. J., Lee, D. J., Krumn, J. T., & Foster, J.E. (2007). Population variation of *Spodoptera frugiperda* (J.E. Smith) in the Western Hemisphere. *Journal of Insect Science*, 7: 1-10.
- Cruz, I., Figueredo, M. L. C., Oliveira, A. C., & Vasconcelos, C. A. (1999). Damage of Spodoptera frugiperda (Smith) in different maize genotypes cultivated in soil under three levels of aluminium saturation. International Journal of Pest Management, 45: 293-296.
- Deshmukh, S., Kalleshwaraswamy, C. M., Asokan, R., Swamy, H. M. M., Maruthi, M. S., Pavithra, B. P., Hegde, K., Navi, S., Prabhu, S. T., & Goergen, G. (2018). First report of the fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae), an alien invasive pest on maize in India. Pest Management in Horticultural *Ecosystems*, 24: 23–29.
- Dumas, P., Legeai, F., Lemaitre, C., Scaon, E., Orsucci, M., Labadie, K., Gimenez, S., Clamens, A. L., Henri, H., Vavre, F., Aury, J. C., Fournier, P., Kergoat, G. J., & Alencon. E. (2015) Spodoptera frugiperda (Lepidoptera: Noctuidae) host-plant variants: Two host strains or two distinct species ? Genetica, 143: 305–316.
- FAO (2020). Global Action for Fall Armyworm Control. Available online: http://www.fao.org/fallarmyworm/fawmanagement/zh/
- Farahani, S., Naseri, B., & Talebi, A. A. (2011). Comparative life table parameters of the beet armyworm, *Spodoptera exigua* (Hübner) (Lepidoptera, Noctuidae) on five host plants. *Journal of the Entomological Research Society*, 13(1): 91-91.
- Goergen, G., Kumar, P. L., Sankung, S. B., Togola, A., & Tamò, M. (2016). First report of outbreaks of the fall armyworm *Spodoptera frugiperda* (JE Smith) (Lepidoptera, Noctuidae), a new alien invasive pest in West and Central Africa. *PloS ONE*, *11*(10): e0165632. doi:10.1371/journal. pone.0165632
- Jing-FeiGuo, Zhang, M. D., Gao, Z. P., Wang, D. J., He, K. L., & Wang, Z. Y. (2021). Comparison of larval performance and oviposition preference of *Spodoptera frugiperda* among three host plants: potential risks to potato and tobacco crops. *Insect Science*, 28: 602–610.
- Johnson, S. J. (1987). Migration and the life history strategy of the fall armyworm, *Spodoptera frugiperda* in the western hemisphere. *International Journal of Tropical Insect Science*, 8: 543-549.
- Luginbill, P. (1928). The fall armyworm. USDA Technical Bulletin, 34:1-94.

Kranthi & Devi

- Li-mei, H. E., Sheng-yuan, Z. H. A. O., Xi-wu, G.A.O & Kong-ming, W.U. (2021). Ovipositional responses of *Spodoptera frugiperda* on host plants provide a basis for using Bt-transgenic maize as trap crop in China. *Journal of Integrative Agriculture*, 20(3): 804-814.
- Marenco, R. J., Foster, R. E., & Sanchez, C. A. (1992). Sweet corn response to fall armyworm (Lepidoptera: Noctuidae) damage during vegetative growth. *Journal* of Economic Entomology, 85: 1285-1292.
- Martinelli, S., Clark, P. L., Zucchi, M. I., Silva-Filho, M. C., Foster, J. E., & Omoto, C. (2007). Genetic structure and molecular variability of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) collected in maize and cotton fields in Brazil. Faculty Publications: Department of Entomology, 164.
- Mitchell, E. R. (1979). Migration by Spodoptera exigua and S. frugiperda, North American style. Movement of highly mobile insects: concepts and methodology in research. North Carolina State University, Raleigh, NC, 386-393.
- Montezano, D. G., Specht, A., Sosa-Gómez, D. R., Roque-Specht, V. F., Sousa-Silva, J. C., Paula-Moraes, S.D., Peterson, J. A., & Hunt, T. E. (2018). Host plants of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in the Americas. *African Entomology*, 26(2): 286-300.
- Murúa, Gabriela & Eduardo Virla (2004). Population parameters of Spodoptera frugiperda (Smith) (Lep.:Noctuidae) fed on corn and two predominant grasess in Tucuman (Argentina). Actazoológica Mexicana, 20(1): 199-210.
- Nagoshi, R. N., & Meagher, R. L. (2004). Behavior and distribution of the two fall armyworm host strains in Florida. *Florida Entomologist*, 87(4): 440-449.
- Nagoshi, R. N. (2009). Can the amount of corn acreage predict fall armyworm (Lepidoptera: Noctuidae) infestation levels in nearby cotton ? Journal of Economic Entomology, 102: 210-218.
- Prowell, D.P., McMichael, M., & Silvain, J. F. (2004). Multilocus genetic analysis of host use, introgression, and speciation in host strains of fall armyworm (Lepidoptera: Noctuidae). Annals of the Entomological Society of America, 97: 1034-1044.
- Ribeiro, L. P., Klock, A. L. S., Nesi, C. N., Luczkievicz, F. R. G., Travi, M. R. L., & Rech, A. F. (2020). Adaptability and Comparative Biology of Fall Armyworm on Maize and Perennial Forage Species and Relation with Chemical-Bromatological Composition. *Neotropical Entomology*, 49(5): 758-767.
- Rosa, A. P. A., Trecha, C. O., Alves, A. C., Garcia, L., & Gonçalves, V. P. (2012). Biology and fertility life table of *Spodoptera frugiperda* (J.E. Smith) in strains of corn. *Arquivos do Instituto Biológico*, 79(1): 39-45.

- Quisenberry, S. S. (1991). Fall armyworm (Lepidoptera: Noctuidae) host strain reproductive compatibility. Florida Entomologist, 74(2): 194-199.
- Sá V.G., Fonseca, B.V., Boregas, K.G. and Waquil, J.M. (2009). Survival and Larval Development of Spodoptera frugiperda (J E Smith) (Lepidoptera: Noctuidae) on Alternatives Host. *Neotropical Entomology*, 38(1):108-115.
- Sharanabasappa., Kalleshwaraswamy, C. M., Maruthi. M. S., & Pavithra, H. B. (2018). Biology of invasive fall army worm *Spodoptera frugiperda* (JE Smith) (Lepidoptera: Noctuidae) on maize. *Indian Journal of Entomology*, 80(3): 540-543.
- Sharma, D. (1994). Biology and food preference of tobacco caterpillar, *Spodoptera litura* Fabricius, on five different hosts. *Journal of Entomological Research*, 18(2):151-155.
- Silva, D. M., de Freitas Bueno, A., Andrade, K., dos Santos Stecca, C., Neves, P. M. O. J., & de Oliveira, M. C. N. (2017). Biology and nutrition of Spodoptera frugiperda (Lepidoptera: Noctuidae) fed on different food sources . *Scientia Agricola*, 74(10) : 18-31.
- Sparks, A. N. (1979). Fall Armyworm Symposium: A review of the biology of the fall armyworm. *Florida Entomologist*, 62(2): 82-87.
- Tinoco-Ojanguren, Rolando & David C. Halperin (1998). Poverty, production, and health: inhibition of erythrocyte cholinesterase via occupational exposure to organophosphate insecticides in Chiapas, Mexico. *Archives of Environmental Health: An International Journal*, 53(1): 29-35.
- Wang, W., Pengyang, He., Zhang,Y., Liu, T., Jing, X., & Zhang, S. (2020). The Population Growth of *Spodoptera frugiperda* on Six Cash Crop Species and Implications for Its Occurrence and Damage Potential in China. *Insects*, 11: 639.
- Westbrook, J., Nagoshi, R., Meagher, R., Fleischer, S., & Jairam, S. (2016). Modeling seasonal migration off all armyworm moths. *International Journal* of *Biometeorology*, 60: 255–267.
- Wijerathna, D. M. I. J., Ranaweera, P. H., Perera, R. N. N., Dissanayake, M. L. M. C. & Kumara, J. B. D. A. P. (2021). Biology and Feeding Preferences of Spodoptera frugiperda (Lepidoptera: Noctuidae) On Maize and Selected Vegetable Crops. The Journal of Agricultural Sciences - Sri Lanka, 16(1): 126-134.
- Williams, T., Goulson, D., Caballero, P., Cisneros, J., Martınez, A. M., Chapman, J. W., & Cave, R. D. (1999). Evaluation of a baculovirus bioinsecticide for small scale maize growers in Latin America. *Biological Control*, 14(2): 67-75.
- Xue-Ming, Yun-Hong Pang, Hong-Ta Wang., Qing-Liang Li & Tong-Xian Liu (2010). Effects of four host plants on biology and food utilization of the cutworm, *Spodoptera litura. Journal of Insect Science*, 10(1): 22.

How to cite this article: Kranthi, P. and Devi, R.S. (2021). Comparative Biology of Fall Armyworm, *Spodoptera frugiperda* on Different Host Plants under Laboratory Conditions. *Biological Forum – An International Journal*, *13*(4): 381-387.